



NUCLEODUR® RP-Säulen

Bitte beachten: Allen HPLC-Säulen von MACHEREY-NAGEL liegt ein Zertifikat bei, dem spezifische Daten und Testergebnisse der Säule entnommen werden können.

Inhaltsübersicht

- Sicherheitshinweise, Beschreibung der Säulen, Installation, Vorsäulen, Probe, Eluent, Flussrate und Druck, Temperatur, Detektion, Säulenaufbewahrung, Behebung möglicher Fehler, Säulenregenerierung, Zusammenfassung

Sicherheitshinweise

Beachten Sie die allgemeinen Gefahrenhinweise für die jeweiligen Mobilphasensysteme (z.B. Acetonitril oder Methanol) und treffen Sie beim Arbeiten entsprechende Schutzmaßnahmen.

Beschreibung der Säulen

Als stationäre Phase enthalten die NUCLEODUR® RP-Säulen eine nach einem speziellen Verfahren modifizierte C18-, C8- oder spezielle RP-Phase auf Basis von voll synthetischem, sphärischem Kieselgel (Typ B).

Table with columns: NUCLEODUR® RP-Phase, Modifizierung, Eigenschaft / Stabilität. Lists various column types like C18 Gravity, C18 Gravity-SB, etc.

Hier nicht aufgeführte NUCLEODUR® RP-Säulen (z.B. neue Entwicklungen) können ebenfalls entsprechend dieser Gebrauchsanweisung angewendet und behandelt werden.

Installation

Der Einbau der HPLC-Säulen sollte unter Berücksichtigung der Flussrichtung, die auf dem Säulenetikett vermerkt ist, erfolgen.

Vorsäulen

Zum Schutz und zur Verlängerung der Lebensdauer der Säule sollten immer Vorsäulen verwendet werden. Die Filterelemente und das Sorbens der Vorsäule halten Verunreinigungen aus der Probe oder dem Eluenten zurück.

Probe

Die Probe wird in der Regel im Eluenten gelöst und vor der Aufgabe auf die Säule durch die Verwendung eines Spritzenvorsatzfilters (z.B. CHROMAFIL® Xtra PET, 0,45 µm, 25 mm, REF 729220) gereinigt.

Eluent

Die RP-Säulen werden mit dem Eluenten Acetonitril – Wasser (je nach Typ 80:20, 70:30 oder 60:40, v/v; siehe Säulenzertifikat) ausgeliefert. Als Eluenten können typische RP-Eluenten (z.B. Acetonitril oder Methanol mit reinem Wasser oder Puffer; Phosphat- und Boratpuffer nur bis pH 9 und bei Raumtemperatur) verwendet werden.

Flussrate und Druck

Die Flussrate (empfohlen für analytische Säulen mit 2–4,6 mm ID: 0,2–2,0 mL/min) beeinflusst den Zeitaufwand der Trennung, die Auflösung und die Lebensdauer der Säule.

Table: Maximaler Druck [bar] vs ID [mm]. Columns for ID 2, 3, 4, 4.6, 8, 10, 16, 21, 32, 40, 50.

Methanol – Wasser Gemische durchlaufen bei ca. 40% Methanolanteil ein Viskositätsmaximum. Änderungen der Eluentenzusammensetzung sollten daher bei niedriger Flussrate durchgeführt werden.

Temperatur

Säulentemperaturen bis zu 60 °C sind für Methanol – Wasser bzw. Acetonitril – Wasser geeignet. Bei Verwendung von Phosphatpuffern sollte die Temperatur nicht höher als 40 °C sein.

Detektion

Mit den Säulen können spektralphotometrische, refraktometrische und elektrochemische Detektoren benutzt werden. NUCLEODUR® C18 Gravity, C8 Gravity, C4 Gravity, C18 Gravity-SB, C18 Isis, C18 Pyramid, PolarTec, Phenyl-Hexyl, PFP, Sphinx RP und C18 HTec eignen sich ebenfalls für die LC/MS-Detektion.

Equilibration

Bevor Proben gemessen werden können, muss die Säule mit dem Eluenten bei gleicher Flussrate und Temperatur der anzuwendenden Methode gespült werden. Die Säule ist equilibriert, wenn die Basislinie des Detektors keine Drift mehr aufweist (i. d. R. nach 10 Säulenvolumina).

Säulenaufbewahrung

Für die Aufbewahrung wird der ursprüngliche Eluent (siehe Eluent) empfohlen. Verwenden Sie für die Langzeitlagerung keine mobilen Phasen, die anorganische Salze enthalten (siehe Regenerierung).

Behebung möglicher Fehler

Das folgende Schema beschreibt typische Symptome eines Leistungsverlustes und deren Ursache. Alle Säulen unterliegen den strengen Richtlinien und Kontrollen unserer Qualitätssicherung.

Table: Symptom / Fehler / Ursache vs Vorbeugung / Behebung. Lists issues like Baseline drift, Broad peaks, Peak overlap, etc.

Säulenregenerierung

In einigen Fällen kann die Trennleistung der Säule wiederhergestellt werden, indem man die Verunreinigungen vom Sorbensbett entfernt bzw. die Phase regeneriert.

- 1. Frischen Eluenten zubereiten: Manchmal wird der Leistungsabfall durch eine Verunreinigung des Eluenten verursacht. ... 2. Reinigen des Sorbens: Zur Entfernung von Verunreinigungen spülen Sie die Säule mit mind. 10 Säulenvolumina ... 3. Regenerierung: Nach der Anwendung von Puffern spülen Sie unmittelbar nach dem Abschluss der Messreihe ... 4. Säulenaustausch: Die hier beschriebenen Vorschläge können die Trennleistung der Säule leider nicht in allen Fällen wieder herstellen.

Table: Säulenvolumen [mL] vs Länge [mm] and ID [mm].

Zusammenfassung

- Um die Lebensdauer der Säule zu verlängern, berücksichtigen Sie bitte folgende Hinweise: 1. Als Eluenten werden organisch-wässrige Eluentensysteme empfohlen (z.B. Acetonitril oder Methanol – Wasser oder Puffer). ... 7. Benutzen Sie für alle Arbeiten Reagenzien von mindestens p.A. Qualität und Lösemittel in HPLC-Qualität.

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NUCLEODUR® RP columns

Note: All HPLC columns from MACHEREY-NAGEL are supplied with a certificate, which contains specifications and test results of the column. NUCLEODUR® RP columns are quality products based on the high purity and very pressure stable silica NUCLEODUR®. They are specifically developed for HPLC analysis. If carefully and properly used excellent chromatographic results and long column lifetime can be achieved. HPLC columns are designed for qualitative and quantitative analysis of mixtures of substances and single components. They must exclusively be used in accordance with universally accepted laboratory regulations and HPLC working methods. Before running the column the entire analytical system (column and equipment) must be carefully checked by the operator. Chromatographic conditions (mobile phase, flow, temperature etc.) has to be adapted to the analytical task. MACHEREY-NAGEL does not give any warranty and is not liable for the success of a separation or application. If you have any questions after reading this manual, please call our service / technical support.

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Safety indication

Follow the general safety instructions for handling of HPLC solvents used as mobile phases (e.g., acetonitrile, methanol) and take precautions against any kind of injuries or damage to health (e.g., skin and eye protection in case of broken capillaries). Disposal of used HPLC columns must follow international, national and local environmental protection regulations. The use of HPLC columns is only permitted to staff members, who are qualified in their field. Keep HPLC columns away from children. MACHEREY-NAGEL disclaims and excludes all warranties of any kind or nature whatsoever and MN shall not be liable for any damages (whether direct, indirect, foreseeable, incidental, compensatory, consequential or special), whether based upon warranty, contract, tort or strict liability, if damages and/or losses occur caused by improper use, maintenance, neglect or improper treatment (especially opening of the column and exposure of the column bed).

Description of the column

As stationary phase NUCLEODUR® RP columns contain a C₁₈, C₈ or special RP phase based on fully synthetical spherical silica (type B), modified with a special method.

NUCLEODUR® RP phase	Modification	Property / Stability
C ₁₈ Gravity	octadecyl, high density, multi-encapsulation	strongly hydrophobic, pH 1–11, LC/MS
C ₁₈ Gravity-SB	octadecyl (monomeric), extensive endcapping	hydrophobic, polar, pH 1–9, LC/MS
C ₈ Gravity	octyl, high density, multi-encapsulation	weakly hydrophobic, pH 1–11, LC/MS
C ₁₈ Isis	octadecyl, specially crosslinked, multi-encapsulation	hydrophobic, weakly polar, steric selectivity, pH 1–10, LC/MS
C ₁₈ Pyramid	octadecyl, polar endcapping	hydrophobic, polar, stable in 100 % aqueous eluents, pH 1–9, LC/MS
PolarTec	RP phase with embedded polar group, endcapping	hydrophobic, polar, steric selectivity, stable in 100 % aqueous eluents, pH 1–9, LC/MS
Phenyl-Hexyl	phenyl-hexyl, multi-encapsulation	hydrophobic, aromatic selectivity, pH 1–10, LC/MS
π ²	biphenylpropyl, multi-encapsulation	hydrophobic, aromatic selectivity, stable in 100 % aqueous eluent, pH 3–10
PFP	pentafluorophenyl, endcapping	hydrophobic, polar, aromatic and steric selectivity, pH 1–9, LC/MS
Sphinx RP	bifunctional octadecyl / phenyl, endcapping	hydrophobic, polar, selectivity for aromatic compounds, pH 1–10, LC/MS
C ₁₈ HTec	octadecyl with high capacity, high density, multi-encapsulation	strongly hydrophobic, pH 1–11, LC/MS
C ₁₈ ec	octadecyl, medium density, endcapping	hydrophobic, pH 1–9
C ₈ ec	octyl, medium density, endcapping	weakly hydrophobic, weakly polar, pH 1–9
C ₄ ec	butyl, medium density, endcapping	weakly hydrophobic, polar, pH 1–9
C ₄ Gravity	butyl, high density, multi-encapsulation	weakly hydrophobic, pH 1–11, LC/MS

NUCLEODUR® RP columns not listed here (e.g., latest developments) can be used and treated in reference to this manual.

Installation

The column should be installed in the flow direction indicated on the column label. It is connected with 1/16" capillaries and fittings, typical for HPLC instruments.

Guard columns

For protection and an extension of column lifetime the column should always be used with a guard column. The filter elements and the adsorbent in the guard column retain contaminants from the sample or the eluent. Connection of the guard column with the separation column is made by a suitable guard column holder (see www.mn-net.com or MN chromatography catalog). Cartridge replacement is required when increased column pressure and/or loss of performance is observed.

Sample

Sample solutions should be passed through a syringe filter (e.g., CHROMAFIL® Xtra PET, 0.45 µm, 25 mm, REF 729220) before entering the column. If injected sample solutions are still turbid even after filtration, the lifetime of the column may be significantly reduced. The sample volume should be as small as possible to achieve an optimal resolution.

Eluent

RP columns are supplied with the eluent acetonitrile – water (depending on the type 80:20, 70:30 or 60:40, v/v; see column certificate for details). As mobile phase typical RP eluents (e.g., acetonitrile or methanol with pure water or buffer; phosphate and borate buffer only up to pH 9 and room temperature) can be used. Eluents should be filtered through a 0.2–0.45 µm membrane filter and degassed. Please consider the pH stability of the used column. Strong acidic or basic conditions can result in dissolution of the column bed or the organic modification. The amount of buffer salts should be kept as low as possible. Note the solubility limit of the buffer in the eluent. The increase of the organic portion can result in precipitation of buffer salts and plugging of the column. Before start of operation with eluent containing a buffer the column should be first preconditioned with a minimum of 10 column volumes acetonitrile – water (25:75, v/v). Always after finishing measurements with buffer-containing eluents the column should be regenerated (see column regeneration). Use of ion pair reagents with phases with embedded polar groups (e.g., NUCLEODUR® PolarTec) can result in unspecific non-reproducible interactions.

Flow rate and pressure

Flow rate (recommended for analytical columns with 2–4.6 mm ID: 0.2–2.0 mL/min) influences the time required, the resolution and the column lifetime. It is limited by the maximum column back pressure, which should not exceed the limits listed in the table below.

Silica	ID [mm]:	Maximum pressure [bar]										
		2	3	4	4.6	8	10	16	21	32	40	50
NUCLEODUR® 1.8 µm (110 Å)	900	800	600	600	-	-	-	-	-	-	-	-
NUCLEODUR® 3, 5, 7 and ≥ 10 µm (110 Å)	600	600	600	600	400	400	400	400	400	400	400	400
NUCLEODUR® 5 µm (300 Å)	450	450	450	450	400	400	400	400	400	400	400	400

In mixtures of methanol and water viscosity reaches a maximum at about 40 % methanol. For this reason a reduced flow rate is recommended, when changing the eluent composition. We recommend controlling back pressure regularly. If a high pressure results from the use of the column at nominal flow rates, this usually indicates that some contaminants have become deposited on the packing material, which must be removed (see troubleshooting).

Temperature

Column temperatures up to 60 °C are possible for methanol – water or acetonitrile – water. For use of phosphate buffer temperature should not be higher than 40 °C. For a long lifetime 30–40 °C is recommended. However, temperature should be at least 30 °C below the boiling temperature of the eluent, in order to ensure proper detection. Variation of the temperature influences retention times and especially the peak shape. Optimum temperatures for successful separations should be determined empirically.

Detection

Spectrophotometers, refractometers and electrochemical detectors can be used with the columns. NUCLEODUR® C₁₈ Gravity, C₈ Gravity, C₄ Gravity, C₁₈ Gravity-SB, C₁₈ Isis, C₁₈ Pyramid, PolarTec, Phenyl-Hexyl, PFP, Sphinx RP and C₁₈ HTec are also suitable for LC/MS detection. If a higher sensitivity is required, post-column derivatizations with an appropriate detector for the reaction product can be used.

Equilibration

Prior to measurement of samples the column must be rinsed with the eluent at the same flow rate and temperature as the method to be applied. Column equilibration is finished, when the baseline of the detector no longer shows a drift (generally after 10 column volumes).

Column storage

The original eluent (see eluent) is recommended for storage. For long-term storage mobile phases containing inorganic salts are not recommended (see regeneration). Methanol is also not recommended for a longer storage, because of a possible impurity with metal ions (e.g., iron(III)). For column storage be sure the end fittings are tightly sealed using column end plugs, because storage without these seals can result in drying of the packing material. Under these circumstances rinse the column with approx. 10 column volumes of the eluent of storage at a flow rate of max. 0.2 mL/min.

Troubleshooting

The following outline describes the symptoms of performance loss and their cause. All columns are subject to the strict regulation and control of our quality assurance system. Columns based on silica are robust and hold their separation efficiency for long periods by correct maintenance and treatment. According to experience, column failures are mostly a result of injection of contaminants to the sorbent bed. The usage of a guard column, as well as an appropriate sample pretreatment will help to minimize these risks.

Use the outline below to help determine the cause of a possible performance loss:

Symptom / Error / Cause	Prevention / Remedy
Baseline drift · insufficient period for equilibration with the eluent · contaminated eluent · temperature	longer or better equilibration use freshly prepared solvents and reagents column temperature control
Broad peaks · mixing and/or diffusion before/behind the column · too large sample volume	keep length and ID of capillaries at a minimum smaller injection volume
Peak interference; too fast elution too fast elution and/or insufficient separation by: · improper column temperature or flow rate · elution power of eluent is too high	optimize concerned parameter optimize eluent system
Increasing back pressure; degradation of the separation performance contamination of sorbent by: · particulate accumulation on frit or sorbent bed from sample, eluent or system · precipitation of buffer salts	prepare fresh eluent; prefilter samples and eluent, use in-line filter / rinse LC system, clean the sorbent check solubility of buffer salts before / remove them by rinsing (see column regeneration)
Insufficient separation; degradation of the separation with regular column pressure contamination with: · fats, oils, lipids from sample (coating of sorbent surface) and other organic substances from improperly prepared eluent or matrices	remove organic substances by sample preparation / clean the sorbent (see column regeneration)
Double peaks (dead volume) · faulty fittings (capillaries, ferrules, nuts) · dissolution of silica by too high pH value of eluent	use "PEEK Fingertight Fittings", REF 718770 or REF 718778 / replace fittings consider pH range of column / replace column

Column regeneration

In some cases the performance of the column can be restored by removing contaminants from the sorbent bed or by regeneration of the phase. It is important, however, to locate the source of contamination before again using the column for the analysis of samples.

- Prepare fresh eluent:** Sometimes the performance loss is traced to eluent contamination. Therefore, prepare fresh eluent and flush all liquid lines before using the column again. The eluent should be filtered through a 0.2–0.45 µm membrane and degassed prior to use.
- Cleaning of sorbent:** To remove contamination rinse the column with a minimum of 10 column volumes (see table below) at the original flow rate and temperature as follows:
 - acetonitrile – water or methanol – water (10:90, v/v) for removal of the buffer
 - 100 % methanol to remove polar organic compounds
 - 100 % acetonitrile to remove medium polar organic compounds (possibly T= 40 °C)
 - 100 % tetrahydrofuran to remove nonpolar organic compounds
 - if necessary, 100 % tetrahydrofuran with inverse flow direction at 1/5 of original flow rate
 - convert column to storage condition using acetonitrile – water (80:20, 70:30 or 60:40, v/v) at original flow rate

An adequate indicator for a clean column is a constant baseline. At constant temperature you should observe less than 2–3 mAU drift during a running time of 5 minutes with an isocratic run.

- Regeneration:** After the usage of buffer, directly after finishing a measurement and always before storage of the column rinse with a minimum of 10 column volumes at the original flow rate and temperature as follows:
 - acetonitrile – water or methanol – water (10:90, v/v) for removal of the buffer
 - increase the organic part in steps of 20 % to the conditions of a new measurement run
 - or gradually increase the part of acetonitrile in steps of 20 % to the storage conditions
- Column replacement:** The above procedures will restore performance only in certain cases. Some organic contaminants are particularly refractory and may not respond to treatment. Also dead volume, due to column compression can generally not be repaired. Under these circumstances, column replacement is necessary. It is highly advisable to locate the cause of the problem before installing a new column.

Length [mm]	ID [mm]:	Column volume [mL]			
		2	3	4	4.6
100		0.30	0.70	1.25	1.65
150		0.45	1.05	1.90	2.50
250		0.80	1.75	3.15	4.15

Abstract

To extend column lifetime, please keep in mind the following:

- As RP eluents organic-aqueous eluent systems (e.g., acetonitrile or methanol – water or buffer) are recommendable. Please consider regeneration after usage of buffers. Eluents should be filtered through a 0.2–0.45 µm membrane and degassed.
- Filter samples through a 0.2–0.45 µm CHROMAFIL® Xtra PET syringe filter before injection.
- Use a guard column for contaminated samples.
- The recommended flow rate for analytical columns (ID 2–4.6 mm) is 0.2–2.0 mL/min.
- Adjust flow rate to keep column pressure below the maximum value of your column.
- Store the column in acetonitrile – water (80:20, 70:30 or 60:40, v/v) after removal of buffer salts.
- Use analytical grade reagents and HPLC grade solvents for all work. Discard any solutions that show evidence of bacterial growth.